1	Conserved Transcriptomic Profiles Underpin Monogamy across
2	Vertebrates
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Materials and Methods

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Sample collection and RNA extraction

Reproductive males of each focal species were sacrificed and brains were rapidly dissected and stored to preserve RNA (species-specific details provided below). All animal care and use practices were approved by the respective institutions. For each species, RNA from three individuals was pooled to create an aggregate sample for transcriptome comparison. The focus of this study is to characterize similarity among species with independent species-level transitions to a monogamous mating system rather than to characterize individual-level variation in gene expression. Pooled samples are reflective of species-level gene expression variation of each species and limit potentially confounding individual variation for species-level comparisons (1, 2). While exploration of individual variation is critical to identify mechanisms underlying differences in behavioral expression, high levels of variation between two pooled samples of conspecifics could obscure more general species-specific gene expression patterns. Note that two pooled replicates per species would not be sufficiently large for estimating within species variance, and the effect of an outlier within a pool of two individuals would be considerable. Therefore, samples were pooled to minimize the effects of individual variation in mating behavior within each species. For all samples, Total RNA was extracted from brains using the TRIzol protocol (Invitrogen) following homogenization of brain tissue. RNA quality and concentration was determined using the Bioanalyzer (Agilent).

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<u>Voles</u>: We used meadow voles (*M. pennsylvanicus*) that were 3rd and 4th generation descendants of those captured near Oshkosh, Wisconsin, USA. In this study, meadow voles were born and raised under a long photoperiod (14:10 h, L: D, lights on at 0700h CST). We weaned the voles at 19 days of age. We then housed them with littermates until 34 days of age. When we separated littermates, we housed them individually in clear

plastic cages (27 x 16.5 x 12.5 cm, I x w x h) and then maintained the males and females in different rooms. For both species, on the day of euthanasia, each male vole was anesthetized with isoflurane and rapidly decapitated. Brains were removed and frozen on dry ice, then stored at -80° C.

Mice: Adult male and female *P. maniculatus* and *P. californicus* were obtained from the Peromyscus Genetic Stock Center (Univ. South Carolina, Columbia, SC). Caging, diet, and bedding were as previously described (3). For each species we created six malefemale pairs. One week after pairing each mouse was lightly anesthetized with isoflurane and rapidly decapitated. Brains were removed and frozen on powdered dry ice.

<u>Birds</u>: Fieldwork was conducted in the Harghita region of Transylvania, Romania, to collect tissue from wild populations of water pipit, *Anthus spinoletta*, and dunnock, *Prunella modularis*, during their breeding season in May-June 2011 (under permit: Ministerial Order from the Rumanian government no. 1470/2011). Using song playback, four water pipits and five dunnocks (adult males) were lured into mist nests. Morphometric data was collected and collated for each bird. Birds were sacrificed by instantaneous decapitation within four minutes of capture to prevent stress-induced changes to circulating testosterone levels and gene expression (Deviche et al., 2010; Van Hout et al., 2010). Whole brains were dissected out, hindbrains were removed and the remaining material was finely chopped and placed in Eppendorf tubes free from DNA, DNase and RNase, and flooded with RNAlater to remove any air bubbles. Samples were stored on ice for between 8 and 12 hours, to allow the RNAlater to permeate the whole tissue (Applied Biosciences protocol, Ambion), before being stored at approximately –17°C for up to 10 days before being frozen to –80°C.

Frogs: We sampled two species of dendrobatid frogs the monogamous *Ranitomeya imitator* and nonmonogmaous *Oophaga pumilio*. *R. imitator* individuals were captive-bred F3s, approximately one year old, from a breeding colony originally collected in Chazuta, San Martin, Peru. Previous research by one of the authors (K. Summers) and his students on this population confirms monogamous behavior. Adult *R. imitator* males were purchased from Understory Enterprises (Chatham, ON, Canada) and were sacrificed upon arrival. Adult individuals of *O. pumilio* were captured in the field by hand on the island of Bastimentos, Bocas del Toro, Panama, and transferred to a breeding colony housed at Tulane University, New Orleans, USA. All individuals used in this study were housed with a female and successfully reared offspring in the captive colony prior to euthanasia and brain collection. Animals were euthanized by rapid cervical transection and brains were rapidly removed and immediately frozen of dry ice.

<u>Fishes</u>: We chose two sister species from the Ectodine cichlid clade of Lake Tanganyika, Africa: *Xenotilapia spiloptera*, a monogamous species with the male and female forming a pair bond and providing parental care, and the closely related *X. ornatipinnis*, a polygynous species in which only the females provide maternal care (6). Adult and sexually mature individuals (with large gonads with distinct and mature sperm packages) were captive-bred F2s, with *X. spiloptera* males engaged in a pair-bond and *X. ornatipinnis* males actively maintaining a territory. Animals were euthanized by rapid cervical transection, brains rapidly removed, and immediately frozen of dry ice.

RNA sequencing and mapping

Following hindbrain removal, RNA was extracted from fore- and midbrain tissue of reproductively active males using Trizol, according to the manufacturer's instructions. An aliquot of total RNA was then run on a Bioanalyzer Nano RNA chip (Agilent) to confirm

RNA integrity was above 7 for each sample. The three individuals within each species were then pooled in equal RNA amounts before extraction of polyadenylated RNA with the Poly A Purist kit (Ambion), according to manufacturer's instructions.

RNA library preparation and sequencing was performed by the Genome Sequencing and Analysis Facility (GSAF) at the University of Texas. Library prep was done using the NEB small RNA kit (cat #E6160L or #E6160S) and sequencing was performed using Illumina HiSeq. Trimmomatic (7) was used to filter and trim reads using the following parameters: -phred33 ILLUMINACLIPadapters.fa:3:30:10 LEADING:3 TRAILING:3 SLIDINGWINDOW:4:15 MINLEN:36, where adapters.fa is a fasta file containing a list of Illumina barcoded adapters. Transcriptomes were assembled *de novo* with Trinity. To reduce redundancy in the assembly, we ran cd-hit-est (parameters: -c 0.98) Reads were mapped to *de novo* assembled transcriptome using the Burrows-Wheeler Alignment tool (bwa-mem, 8). Raw reads for each gene were calculated as the sum of transcript counts obtained using SAMtools (9). Raw reads were normalized as reads per million (RPM).

Orthology inference

Within species pairs, contigs were annotated and gene orthology was defined by aligning assembled contigs and protein sequences of closely related reference genomes (mammals: *Rattus norvegicus*; birds: *Gallus gallus*; frogs: *Xenopus tropicalus*; fishes: *Oreochromis niloticus*) using BLAST (e-value = 1e-5). Genes that reciprocally BLAST to the clade-specific reference genome in both the monogamous and nonmonogamous species were called orthologous (number of orthologous genes in each clade: 11,051 voles; 10,519 mice; 6993 birds; 7035 frogs; 13,135 fishes).

Clade specific gene loss and duplication events obscure the evolutionary history of genes. such that in many cases gene families will contain distinct numbers of paralogs in different clades. One solution is to focus on orthologous gene groups. We tested similarity in transcriptomic profiles of monogamous species across divergent vertebrate clades using orthologous gene group (OGG) expression patterns. OGGs were identified using the sequence based ortholog calling software package OrthoMCL (10). Protein sequences of the reference genomes (listed above) were organized into orthologous gene groups based on sequence similarity. For each reference genome, genes were grouped into orthologous gene groups (as paralogs) when sequence similarity was higher among genes within species than between species. This approach of ortholog calling improves substantially on reciprocal best BLAST hits, which results in loss of up to 60% of true orthologous relationships as it eliminates all paralogous genes (11). Alternatively, databases of OGGs (12, 13) are most useful for traditional model systems with well-sequenced and annotated genomes. Our method of ortholog calling by sequence similarity of target species using OrthoMCL (10) identified 1979 OGGs, while only 355 OGGs were identified using a database approach (eggNOG). While the remaining analysis characterizing monogamyrelated gene-expression patterns focused on OrthoMCL OGGs, similar results were found when eggNOG OGGs were analyzed. Because our study focused on identifying monogamy-related expression patterns, and genes in the same OGG were generally concordant in directionality of expression differences (Fig. S2; Table S3), when an OGG contained more than one gene (i.e., paralogs: Fig. S2; voles: 573, 29.0%; mice: 521, 28.3%; birds: 320, 16.2%; frogs: 227, 11.5%; fishes: 730, 36.9%) the gene with the highest log₂ fold-difference between the monogamous and nonmonogamous species pairs was used for the remainder of the analysis. Similar overall patterns were obtained using the mean log₂ fold-differences for the orthologous gene groups.

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Differential Expression Analysis

To assess concordance of OGG expression in monogamous species across clades we used the differential expression analysis software package DESeq2 (14). For each species, raw read counts for the 1979 OGGs identified across all species were included in the differential expression analysis. Species were labeled as monogamous or nonmonogamous such that species from distinct clades served as biological replicates in the analysis. DESeq2 was performed on different evolutionary sub-groups including mammals, amniotes, tetrapods, and all clades. OGGs with an expression difference of +/- 1 log₂ fold-difference and p-value < 0.1 were characterized as differentially expressed. This fairly liberal cut off captures genes that are generally concordant in expression direction across clades that may otherwise be eliminated.

To extract patterns of gene expression shared among monogamous species across clades, we utilized a hypergeometric approach, the R package Rank-Rank Hypergeometric Overlap (RRHO) (15). In RRHO analysis, rank \log_2 fold-differences in monogamous vs. nonmonogamous mRNA levels are binned into steps (we used the default settings which binned our 1979 OGGs into 45 steps). Each step can be set as a threshold making a continuous threshold scale of differential expression unique to each clade. At each pairwise \log_2 fold-difference threshold (binned OGG step) enrichment of rank correlations between clades is determined using a hypergeometric distribution. RRHO corrects p-values for multiple comparisons using the B-Y procedure (16). Overall significance of rank correlations for each pairwise comparison of clades was calculated using permutation analysis (17).

Novel candidates and GO analysis

To identify the most robust candidate genes associated with a monogamous mating system across species, we combined the results of our differential gene expression analysis with the RRHO analysis (Fig. 4). Specifically, we identified genes that were concordantly differentially expressed (at +/- 1 log₂ fold-difference) between monogamous and nonmonogamous species in four of the five clades, and equivalently identified in at least five of the ten RRHO comparisons which allows one clade to lack concordant expression (Fig. 5; Dataset S1). To capture genes that are generally concordant in expression direction across clades, which may not be captured by threshold differential expression approaches, we used the more liberal expression cut off of +/- 1 log₂ folddifference. Notably, these candidate genes have known roles in synaptic transmission, neuroplasticity, and neurological function among other functions possibly reflecting similarities among monogamous species in the mechanisms through which the brain reward circuitry becomes associated with social affiliation (e.g., reproductive and parental behavior). Several candidate genes are critical for neuronal development as well as synaptic function and plasticity, including: Low Density Lipoprotein Receptor-Related Protein 6 (LRP6) (18), the Wnt inhibitor Adenomatosis Polyposis Coli Down-Regulated 1 (APCDD1) (19), the Lysophosphatidic Acid Receptor 1 (LPAR1) (20), and Notch1 (21). Relatedly, the candidate gene (Dscam) remodels microcircuitry through regulation of dendritic arborization (22), and in this way is also thought to enhance learning and memory (27). Two genes on the candidate list have documented roles in synaptic transmission. The excitatory neurotransmitter Metabotropic Glutamate Receptor 6 (GRM6) is the only known metabotropic glutamate receptor to directly mediate synaptic transmission in the nervous system (23). Huntingtin Interacting Protein (Hip1) regulates AMPA receptor trafficking (24) and, interestingly, also enhances androgen receptor-mediated transcription (25). Finally, candidate genes lysine methyltransferase 2C (KMT2c) (26) and solute carrier family 6 member 17 (SLC6a17) (27) are involved in cognitive function and cause

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intellectual disability when mutated. Note that in mice, the candidate genes Tnik, Lpar1, Man2a1, and Lrp6 predominantly show expression in the forebrain, whereas the remaining genes are either broadly distributed throughout the brain (28) or their brain expression has yet to be mapped.

To characterize similarity of function of monogamy-related OGGs across species we assessed concordance of over- and under-represented GO annotations using BiNGO (29). For each clade, enrichment of GO terms was assessed for OGGs up- or down-regulated at log_2 fold-difference +/- 1 with the complete list of 1979 OGGs as the reference set. BiNGO uses a parent-child approach taking into account dependencies among GO terms. GO terms were identified as over- or under-represented using a hypergeometric test. p-values were then adjusted for multiple comparisons using Benjamini & Hochberg's FDR correction.

Phylogenetic, life history, and gene expression distances

As described above, we chose species pairs with similar ecological attributes (except for mating system characteristics) for each clade to control for the potential confounding effects of a species' natural history. For each species, we consulted the literature to score characteristics of mating system (i.e., presence of a pair bond, territoriality, and direct and indirect paternal care) and ecology (i.e., habitat complexity, activity patterns, diet, and gregariousness) (SI Appendix, Tables S1 and S2).

To assess the contribution of evolutionary history and mating system on neural gene expression, we compared evolutionary distance, and similarities in mating system characteristics to gene expression distance between all pairs (Fig. 6A and B). A mating system score was calculated for each species as the sum of the mating system

characteristics, where higher values indicate more elaborated monogamy (i.e., males consistently form pair bonds, provide both direct and indirect parental care, exhibit high levels of territoriality, and are less sexually dimorphic). We estimated evolutionary divergence between clades using TimeTree (30). For each clade, expression distance was estimated as the Euclidean distance in log₂ fold-difference of expression between the monogamous and nonmonogamous species (31). To remove the variation in gene expression and mating system characteristics due to phylogeny, we calculated phylogenetic independent contrast on the first principal component (PC) of gene expression and mating system score using the R package ape (32). In both the Euclidean distance comparisons (Fig. 6A and B) and the principal components analysis (PCA) (Fig. 6C), we limited the OGGs to include only those that were differentially expressed (±1 log₂ fold difference in at least one clade) and variable (upper quartile of variance across all species). This subset included 401 OGGs that overlap significantly with candidate OGGs discovered using RRHO (overlap = 35, hypergeometric distribution p = 7.6e-07) and differential expression analysis (overlap = 83, hypergeometric distribution p = 3.7e-13) (Fig. S7). A PCA including log₂-transformed RPM for this subset of OGGs in all 10 species was performed using the prcomp function in R. PC1 described 22.8% of the variation in expression.

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Accessibility of Data and Data Analyses

RNA sequencing data discussed in this publication have been deposited in NCBI's Gene Expression Omnibus (33) and are accessible through GEO Series accession number GSE123301 (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE123301). Additional data and data analyses scripts in R and Python are published and publically available as a dataverse at the Texas Data Repository (https://dataverse.tdl.org/dataverse/monogamy).

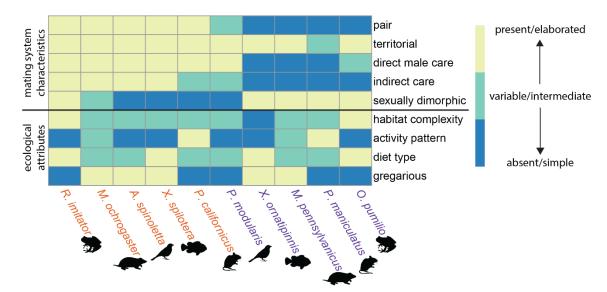


Figure S1. Ecological attributes and mating system characteristics of study species. Species names in orange are the monogamous species, those in purple are the nonmonogamous species for each clade. Color indicates presence or degree of trait elaboration in that species unless otherwise indicated below. Yellow indicates the trait listed on the right is absent or 'simple', green indicates intermediate, and blue indicates present or 'elaborated'. Five mating system characteristics were scored, including: pair bond formation, defense of breeding territories, male engagement in offspring provisioning, cleaning, or transport, males actively defend nests or provision females, and presence of sexual dimorphism (size or color elaboration). Four additional ecological attributes were also scored, including: habitat complexity (simple = yellow, various/intermediate = green, or complex environments = blue), activity pattern (diurnal = yellow, crepuscular/seasonally variable = green, and nocturnal = blue), diet type (primary diet is "herbivorous" = yellow, "omnivorous" = green, and "carnivorous" = blue), and whether communal or gregarious behaviors are observed outside of a reproductive context. Details and references in (SI Appendix, Tables S2 and S3).

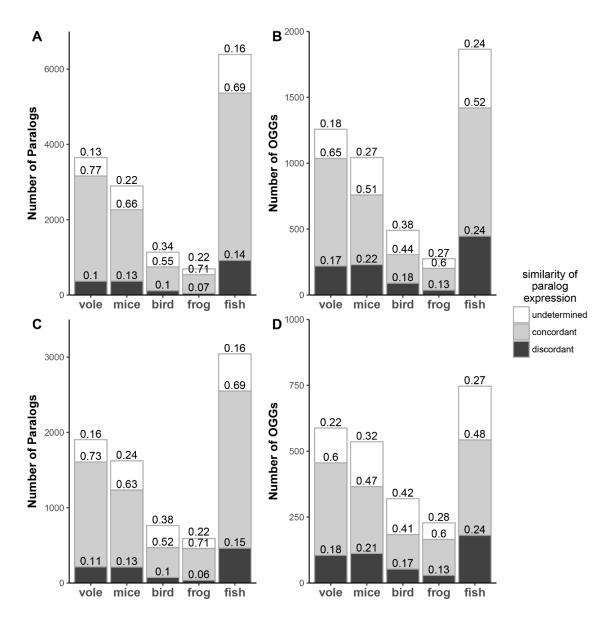


Figure S2. To compare neural gene expression across all clades, genes were grouped into orthologous gene groups (OGGs) using orthoMCL. OrthoMCL identified 6125 OGGs shared across the rat, chicken, frog, and tilapia reference genomes. Of those, 44-76% were identified using RNAseq (Table S3). Due to variation in paralog identification, different numbers of genes were included for each clade (voles: 7540, mice: 7046, birds: 4914, frogs: 3703, fish: 10154) (Table S3). 1979 OGGs were shared across all clades with different numbers of paralogs in each clade (Table S4). We assess the concordance in paralog expression at the gene (A and C) and OGG (B and D) levels for all clade-specific

OGGs (A and B) and for OGGs shared across clades (C and D). Paralogs are generally concordant in expression direction between the monogamous and nonmonogamous species (i.e. higher or lower expression). The paralog with the largest fold-difference in expression between the monogamous and nonmonogamous species pairs was selected as the representative gene for each OGG. If this value was < +/- 1 log₂ fold-difference the OGG and its containing paralogs were labeled at "undetermined" (white; all plots). The number of paralogs directionally discordant from the representative gene are shown for each clades (black; A and C). Any OGG containing a discordant paralog is considered discordant (black; B and D). The number of concordant paralogs and OGGs are shown in grey. Values at the boundaries indicate the proportion of paralogs (A and C) or OGGs containing paralogs (B and D).

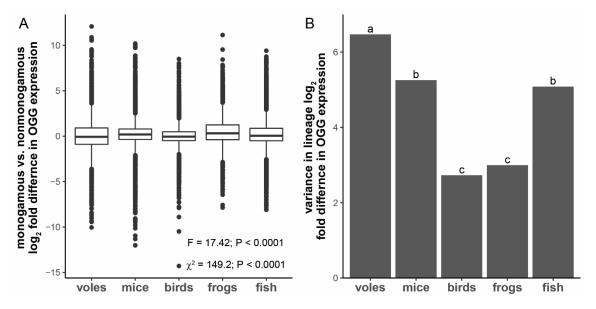


Figure S3. \log_2 fold difference (monogamous vs. nonmonogamous) in orthologous gene group (OGG) expression for all 1979 orthologous gene groups in each clade (A). Variance in \log_2 fold-difference (monogamous vs. nonmonogamous) in OGG expression (B). When more than one gene is present in an orthologous gene group the gene with the highest \log_2 fold-difference was selected. \log_2 fold-difference in OGG expression in monogamous species is slightly skewed toward increased expression in mice and frogs (median = 0.19 and 0.31, respectively). Birds and frogs exhibit the smallest variance (B) and narrowest interquartile range of expression differences (A) between monogamous and nonmonogamous species. Overall, means (Kruskall-Wallis \square^2) and variances (Levene's test) differ among the clades. F-tests for equality of variances were used to compare variances for all pairwise clades. Clades separated by letters significantly differ after correcting p values for multiple hypothesis testing (p * number of comparisons).

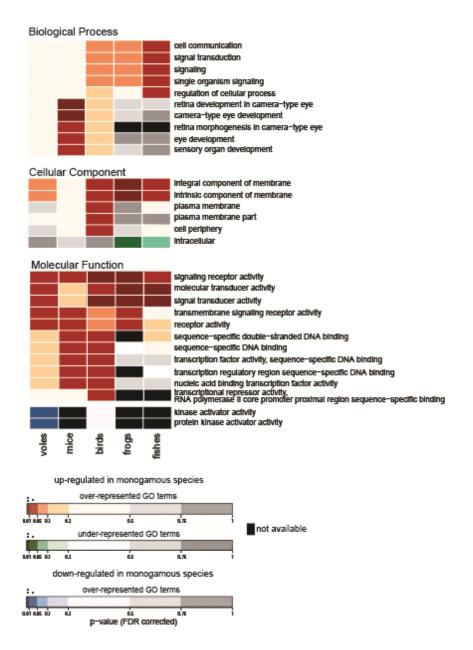


Figure S4. To characterize similarity of function of monogamy-related orthologous gene groups (OGGs) across species we assessed concordance of over- and under-represented GO annotations using BiNGO (29). For each clade, enrichment of GO terms was assessed for OGGs up- or down-regulated at log₂ fold-difference +/- 1 with the complete list of 1979 OGGs as the reference set.

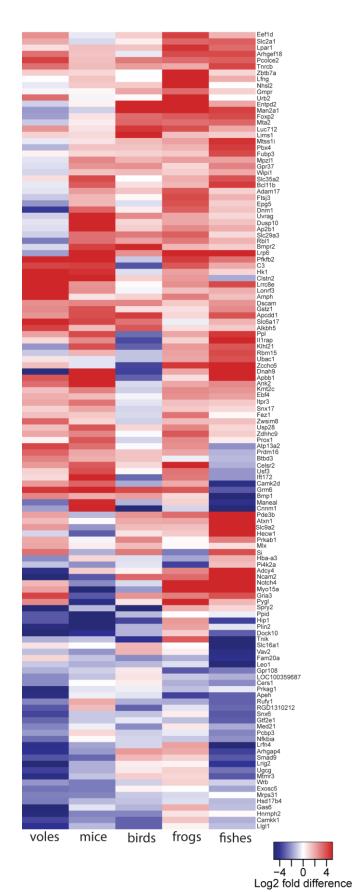


Figure S5. Genes concordantly expressed in monogamous brains across vertebrates. 123 genes were identified with DESeq2 log₂ fold-difference +/- 1 and p-value < 0.1. DESeq2 provides a base mean and a log₂ fold-difference. As with any other analysis with replicates, not all replicates are expected to show the same directional difference in gene expression. However, on average, these 123 OGGs are differentially expressed between monogamous and nonmonogamous species across all clades. Gene symbols are provided where available, when gene symbols are not available the Ensembl rat protein id is provided.

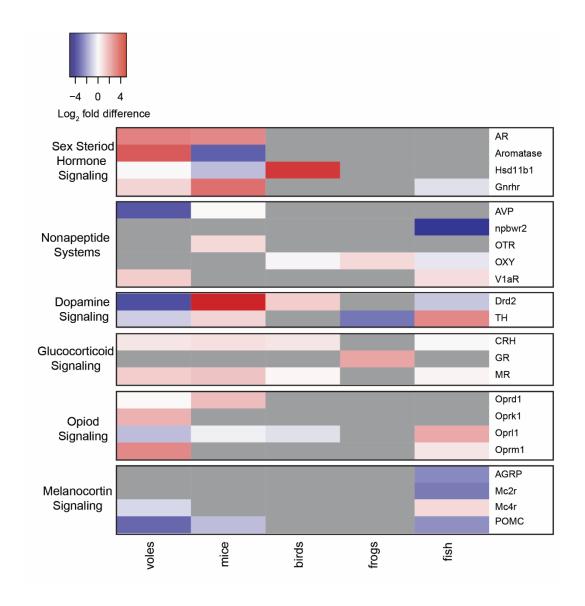


Figure S6. Relative expression (log₂ fold-difference) in monogamous versus nonmonogamous species of each clade for 22 candidate genes previously implicated in regulating complex social behavior across vertebrates. These often studied candidate genes represent six neuroendocrine and neuro-modulatory systems that have previously implicated in the regulation of (aspects of) monogamous behavior or, more generally, complex social behavior. Reds (from light to dark) indicate increased expression in the monogamous species; blues (from light to dark) indicate decreased expression in the monogamous species. Grey indicates that expression data was not available for one or both species of that clade. Note that in several clades many of these candidate genes

- were not detected in either the RNAseq analysis or during the orthology inference step,
- 319 therefore limiting the interpretability of these data.

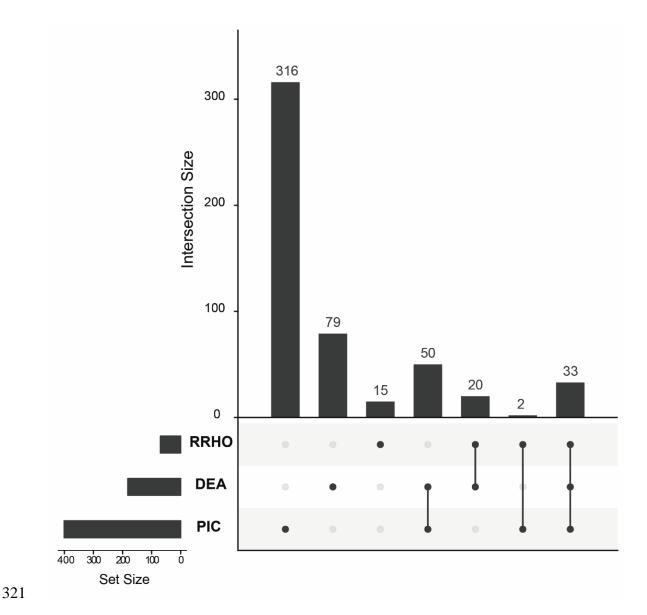


Figure S7. 401 OGGs exhibit both differential expression between the monogamous and nonmonogamous species in at least one clade (at +/- 1 log2 fold difference) and high expression (RPM) variance across all species (variance in the upper quartile). This subset included 401 OGGs used in the phylogenetic independent contrast (PIC) overlap significantly with 70 OGGs identified by RRHO (overlap = 35, hypergeometric distribution p = 7.6e-07) and 182 OGGs identified using differential expression analysis (overlap = 83, hypergeometric distribution p = 3.7e-13). Intersections were identified and plotted using the R package UpSetR (34). P-values are corrected for multiple hypothesis

testing. RRHO candidate OGGs are among the most up- or the most- down-regulated in 6 of the 10 comparisons (as in Fig. 5). DEA candidate OGGs include those identified with DESeq2 across all vertebrates (\log_2 fold-difference +/- 1 and p-value < 0.1) or those that exhibit a +/- 1 \log_2 fold expression difference between the monogamous and nonmonogamous species in at least four clades (as in Fig. S5 and Fig. 5, respectively).

Table S1. Mating system characteristics of each species used.

species	pair bond	territorial	paternal offspring care	indirect paternal care	sexually dimorphic
M. ochrogaster	Yes (35, 36)	Yes (36, 37)	Yes (38)(39)	Yes (40, 41)	Various (3, No: 8, Yes: in the lab: 9)
M. pennsylvanicus	No (44)	Yes (45, 46)	No (41)	No (41)	Yes (47)
P. californicus	Yes (48, 49)	Yes: highly aggressive and territorial (50, overlapping territories: , 51)	Yes (52–54)	Yes (55–58)	No (most sources say no;but, brain regions are sexually dimorphic 59)
P. maniculatus	No (60, 61)	Yes: much less so than P. californicus (62, 63)	No (64, 65) (note: pup licking/nest sitting reported in one study)	No (no reports)	Yes (43)
A. spinoletta	Variable (86%: 66)	Yes (66)	Yes (67)	Yes: alarm calling, feeding & incubating females (66)	No (66)
P. modularis	Variable depending on space use (68, 69)	Yes: mate guarding (70)	Yes: provisioning (70)	Sometimes (71)	No (72)
R. imitator	Yes (73, 74)	Yes (73)	Yes: egg attendance; tadpole feeding and transport (75)	Yes (73)	Yes: (females are slightly, but significantly larger 73)
O. pumilio	No (76, 77)	Yes (76, 78)	Yes: intermediate: periodic water shedding (79)	No (80)	Yes: (females larger than males, C. Richards-Zawacki pers. obs.)
X. spilotera	Yes (81)	Yes (81)	Yes (81, 82)	Yes (81)	No (HAH pers. obs.)
X. ornatipinnis	No (82)	Yes: 'roving territories' (HAH pers. obs.)	No (82)	No (82)	Yes: chin pigmentation in males (HAH pers.l obs.)

Table S2. Ecological attributes of the study species.

species	habitat complexity	activity	diet	communal/gregarious outside of breeding
M. ochrogaster	Intermediate: un- grazed grassland/savanna (37)	Various, seasonal (83)	Omnivorous: forbs, foliage, fruits, tubers, insects (84, 85)	Yes: communal group living (86, 87)
M. pennsylvanicus	Intermediate: grasslands, woodlands, riparian (88)	Various depending on landscape and season (89)	Omnivorous: grasses, foliage, fruits, tubers, insects (89, 90)	Yes: communal nesting in winter populations (91, 92)
P. californicus	Intermediate: dense chaparral/broad- sclerophyll woodland (48)	Nocturnal (93)	Omnivorous: acorns, flowers seeds, fungi and arthropods (94)	No: non-overlapping territories (51)
P. maniculatus	Various (95)	Nocturnal (96)	Omnivorous: mostly insects & arthropods, but also seeds, flowers, and leaves (63, 97)	No (62, 63)
A. spinoletta	Intermediate: open, heterogeneous habitats: shrub lands, medium woodlands, wet inlands (98, 99)	Diurnal	Omnivorous: mostly insects and seeds (100)	Yes: feeds in large groups outside of the breeding season (101)
P. modularis	Intermediate: woodlands, hedgerows, gardens	Diurnal	Omnivorous: mostly insects and seeds (102)	No: solitary in winter (103)
R. imitator	Complex: leaf litter, secondary to old growth forest (73)	Diurnal	Carnivorous: ants, mites, beetles (A. Stuckert, pers. obs.)	No (K. Summers pers. obs.)
O. pumilio	Complex: premontane forest leaf litter	Diurnal (104)	Carnivorous: ants, mites, beetles, springtails(105, 106)	No; (76–78)
X. spilotera	Intermediate (82, 107)	Diurnal (HAH pers. obs.)	Carnivorous (108)	Yes (82, 108)
X. ornatipinnis	Simple, sand (82)	Diurnal (HAH pers. obs.)	Carnivorous (108)	Yes (82, 108)

Table S3. For each species, the number of genes and orthologous gene groups (OGGs) are shown. For each clade the number of shared genes and OGGs are shown. For each evolutionary group (i.e., mammals, amniotes, tetrapods, and vertebrates) the number of shared OGGs is shown. The starting set is limited to 6125 orthologous OGGs identified by orthoMCL as containing genes from each of references genome used in this study (i.e., rat, chicken, *Xenopus*, and tilapia).

Species	Genes	OGGs	clade genes	clade OGGs	mammal OGGs	amniote OGGs	tetrapod OGGs	vertebrate OGGs			
M. ochrogaster	9733	6007	7840	5449 (0.890)							
M. pennsylvanicus	8088	5555	7040		(0.890)	´ 4746					
P. californicus	7936	5578	7046	7046 5193 (0.839) 4914 4267 (0.697)	5193		(0.775) 5193	(0.775)	3508		
P. manipulatus	8298	5679				(0.573)	2081 (0.340)	1979 (0.323)			
A. spinoletta	5378	4914	AU1A -								
P. modularis	6441	5378									
R. imitator	6170	5035	3703	3703	3284						
O. pumilio	4895	4098	0.00	(0.536)							
X. spilotera	11296	5889	10154	5628							
X. ornatipinnis	11401	5882	.5101	(0.919)							

Table S4. For the 1979 orthologous gene groups (OGGs) shared among the clades compared, we show the total numbers and proportions of genes, genes with paralogs, and paralogs with directionally concordant, discordant, and undetermined expression for each clade. In addition, we show the number and proportion of OGGs containing paralogs and OGGs containing directionally concordant, discordant, undetermined paralogs. The paralog with the largest fold-difference in expression between the monogamous and nonmonogamous species pairs was selected as the representative gene for each OGG. If this value was < +/- 1 log₂ fold-difference the OGG and its containing paralogs were labeled as "undetermined."

Clade		total paralogs	Paralogs (proportion of paralogs)		OGGs with paralogs	OGGs (proportion of OGGs with paralogs)			
	genes	(proportio n of genes)	concord ant	discord ant	undeter mined	(proportion of OGGs)	concord ant	discord ant	undeter mined
voles	3294	1903 (0.58)	1392 (0.73)	215 (0.11)	296 (0.16)	588 (0.30)	351 (0.60)	105 (0.18)	132 (0.22)
mice	3066	1623 (0.53)	1023 (0.63)	212 (0.13)	388 (0.24)	536 (0.27)	254 (0.47)	112 (0.21)	170 (0.32)
birds	2421	762 (0.31)	395 (0.52)	74 (0.10)	293 (0.38)	320 (0.16)	131 (0.41)	53 (0.17)	136 (0.43)
frogs	2341	590 (0.25)	421 (0.71)	37 (0.06)	132 (0.22)	228 (0.12)	136 (0.60)	29 (0.13)	63 (0.28)
fish	4275	3043 (0.71)	2087 (0.69)	463 (0.15)	493 (0.16)	747 (0.38)	362 (0.48)	181 (0.24)	204 (0.27)

Table S5. For each clade, taxon median divergence time estimates and references are reported from the TimeTree Database (109). No data is available in the TimeTree Database for *Xenotilapia spilotera*. References used to estimate divergence time between the *Xenotilapia* species are provided.

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Taxon A	Taxon B	divergence time median (MYA)	References:
M. pennsylvanicus	M. ochrogaster	10.5	(110–113)
P. maniculatus	P. californicus	11.5	(110, 111, 113, 114)
Peromyscus	Microtus	18.9	(110, 111, 113, 115–120)
P. modularis	A. spinoletta	29	(121–127)
O. pumilio	R. imitator	34.2	(128, 129)
X. ornatipinnis	X. spilotera	2.5	(6, 130)

Table S6. Maximum, mean, and median –Log₁₀ p-values for each quadrant of the Rank Rank Hypergeometic Overlap analysis (Fig. 4).

		Concor	dant	Discordant				
clade A	clade B	Downs	Ups	Down,Up	Up,Down			
		Max <i>P</i> values						
voles	mice	35.41	6.21	15.70	6.61			
voles	birds	1.71	0.00	0.00	0.00			
voles	frogs	22.33	18.05	20.06	12.35			
voles	fish	22.54	3.40	8.33	6.07			
mice	birds	7.52	10.02	0.27	5.23			
mice	frogs	14.84	14.17	14.84	13.36			
mice	fish	16.95	4.98	9.93	9.55			
birds	frogs	7.91	2.22	8.11	0.00			
birds	fish	5.81	5.43	0.21	0.00			
frogs	fish	16.72	6.42	8.07	14.46			
			Mean F	values				
voles	mice	12.10	0.18	1.52	0.35			
voles	birds	0.17	0.00	0.00	0.00			
voles	frogs	10.48	2.24	6.51	0.94			
voles	fish	8.36	0.16	0.39	0.35			
mice	birds	1.88	0.51	0.00	0.22			
mice	frogs	5.27	0.84	3.18	0.83			
mice	fish	6.52	0.26	0.45	0.86			
birds	frogs	2.54	0.09	2.38	0.00			
birds	fish	1.10	0.59	0.00	0.00			
frogs	fish	7.14	0.28	0.26	3.82			
			Median	<u>P</u> values				
voles	mice	11.51	0.00	0.00	0.00			
voles	birds	0.00	0.00	0.00	0.00			
voles	frogs	11.27	0.00	4.89	0.00			
voles	fish	7.67	0.00	0.00	0.00			
mice	birds	0.93	0.00	0.00	0.00			
mice	frogs	5.38	0.00	0.00	0.00			
mice	fish	6.02	0.00	0.00	0.00			
birds	frogs	2.29	0.00	1.28	0.00			
birds	fish	0.01	0.00	0.00	0.00			
frogs	fish	7.29	0.00	0.00	3.31			

Table S7. Expression of novel candidate genes in monogamous vs. nonmonogamous species pairs.

0	Name -	Duntain ID	log ₂ fold-difference in express				sion
Symbol	Name	Protein ID	voles	mice	birds	frogs	fish
Ank2	Ankyrin 2	ENSRNOP00000015386	4.09	10.01	1.70	2.90	4.52
Apcdd1	Adenomatosis Polyposis Coli Down- Regulated 1	ENSRNOP00000059242	2.82	2.01	4.53	0.28	3.99
Dscam	Down Syndrome Cell Adhesion Molecule (cell adhesion, PAK pathway)	ENSRNOP00000022476	5.45	2.19	2.06	-0.49	1.44
NA	ENSRNOP0000013363	ENSRNOP00000013363	1.48	8.34	1.22	1.35	1.64
ENTPD2	Ectonucleoside triphosphate diphosphohydrolase 2	ENSRNOP0000018560	1.37	1.01	5.26	5.27	2.43
Fam20a	Golgi associated secretory pathway pseudokinase	ENSRNOP00000005367	1.99	-2.83	-2.16	-2.27	-4.85
Grm6	Glutamate Receptor, Metabotropic 6	ENSRNOP00000000249	7.10	5.86	5.26	3.17	2.96
Gtf2e1	General Transcription Factor IIE Subunit 1	ENSRNOP00000039601	-1.12	0.23	-1.32	-1.97	-1.55
Hecw1	HECT, C2 and WW domain containing E3 ubiquitin protein ligase 1	ENSRNOP00000021703	1.87	-2.90	1.38	1.53	7.70
Hip1	Huntingtin interacting protein 1	ENSRNOP00000031153	-2.71	-7.67	-3.54	1.64	-4.70
Kmt2c	Lysine methyltransferase 2C	ENSRNOP00000063937	2.76	2.17	-0.39	2.96	2.55
Lpar1	Lysophosphatidic Acid Receptor 1	ENSRNOP00000043652	3.30	1.77	1.45	2.90	2.72
Lrp6	Low Density Lipoprotein Receptor-Related Protein 6	ENSRNOP00000063261	3.25	6.30	2.26	7.21	5.89
Lrrc8e	Leucine rich repeat containing 8 VRAC subunit E	ENSRNOP00000037487	10.54	2.71	0.23	1.82	4.15
Man2a1	Mannosidase, alpha, class 2A, member 1	ENSRNOP00000020767	1.93	-1.25	4.46	5.72	4.38
Mpzl1	Myelin protein zero-like 1	ENSRNOP00000004376	-0.80	2.45	3.19	1.49	2.18
Notch1	Notch 1	ENSRNOP00000026212	4.01	2.02	-1.42	6.51	7.40
PCOLCE2	Procollagen C-endopeptidase enhancer 2	ENSRNOP00000067519	6.40	0.41	3.16	2.59	3.69
Rbl1	RB transcriptional corepressor like 1	ENSRNOP00000063017	-0.75	1.50	2.23	2.29	2.53
Slc29a3	Solute carrier family 29 member 3	ENSRNOP00000000689	1.88	1.95	3.11	2.91	0.30
Slc6a17	Solute carrier family 6 member 17	ENSRNOP00000065179	9.55	3.58	4.83	-2.68	1.37
SMAD9	Mothers against decapentaplegic homolog 9	ENSRNOP00000000102	-2.70	-2.04	-2.01	0.30	-3.20
Tnik	TRAF2 and NCK interacting kinase	ENSRNOP00000016799	-1.73	-3.35	-4.10	3.00	-5.80
Tnrc6b	Trinucleotide Repeat Containing 6B	ENSRNOP00000049430	5.15	-1.43	3.90	3.51	5.94

Dataset S1. Novel candidate genes associated with monogamous mating system across species. Gene functions are provided by Gene Cards (www.genecards.org) unless otherwise noted. Localizations are provided by the Allen Mouse Brain Atlas (28).

Dataset S2. For each orthologous gene group (OGG) and each clade the gene with the largest expression difference between the monogamous and nonmonogamous species is selected as the representative gene. Ensembl IDs are provided for each OGG and clade in monogamous to nonmonogamous log₂ fold-difference.

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