

Acoustic Monitoring of Estuarine Communities Facing Ecosystem Change ;hull||||he

# **TABLE OF CONTENTS**

## **LISTEN IN**

#### Acoustic Monitoring of Estuarine Communities Facing

Ecosystem Change

Overview	4
Sources of Sound	5
Sound Measurement	5
Data Collections and Analysis	6

#### System Wide Monitoring & Habitat Assessment

The System Wide Monitoring Program of the NERRS	10
SWMP Stations	11
Oyster Reefs	11

#### Stewardship: Impacts of Noise and Monitoring Visitor Use

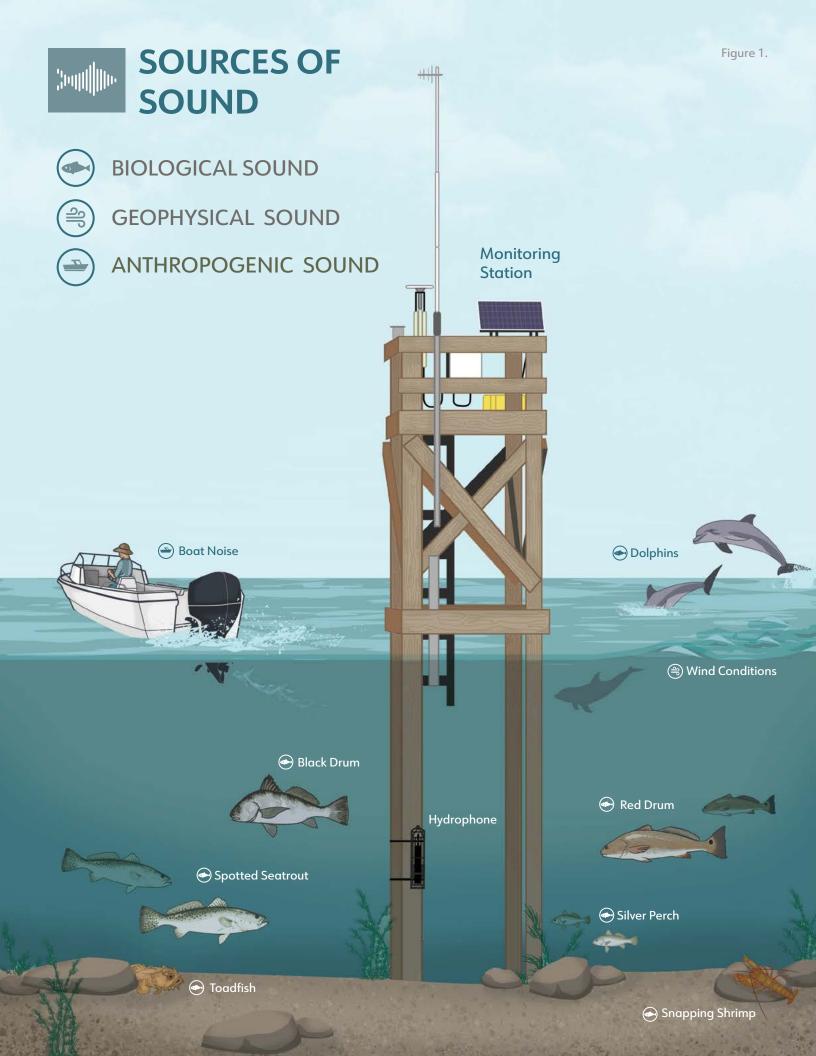
Impacts of Anthropogenic Noise	13
Monitoring Visitor Use	14

#### **Education: Teachers on the Estuary (TOTE)**

Teacher Training Workshop	15
Workshop Activity	16
Student Activities	17

#### References and Resources

References	21
Additional Resources	22
Authorship, Design and Citation	23



## **ACOUSTIC MONITORING** of Estuarine Communities Facing Ecosystem Change

## Overview

'¦uu||**||||**||u

Acoustic monitoring provides time-series data with a high temporal resolution to understand short-term variability and long-term change in aquatic ecosystems. Passive listening devices (hydrophones) record sounds at multiple levels of biological complexity which can be used to investigate and monitor biodiversity, habitat utilization, species distributions, behaviors such as feeding and spawning, phenology, and anthropogenic noise <sup>1, 2, 3,4,5</sup>. By monitoring these parameters, soundscape ecology has the potential to provide insight into the response and resilience of ecosystems, habitats, and individual species to rapidly changing environmental parameters, climate change, and human ocean use<sup>6,7,8</sup>. The products and outputs of this research and data collection will inform management decisions regarding fisheries productivity, habitat restoration and rates of restoration of ecosystem function, while building a baseline of acoustic activity associated with the timing of important phenological events, such as spawning seasons.

By combining acoustic data with traditional environmental monitoring, scientists and managers can identify key habitats for protection and measure how ecological communities respond to environmental changes (e.g. storm events, coastal development, eutrophication) in a cost-effective and low-impact manner<sup>9,10</sup>. In addition to providing information on biodiversity and population status, acoustic monitoring also provides information on ecological processes and habitat status. For example, baseline acoustic data can help to identify deviations in behavior, biodiversity or habitat use due to stochastic events or changing climate patterns/environmental variables<sup>4,6</sup>. The monitoring and research framework outlined below aims to catalyze a multi-sector regional collaboration that can leverage expertise and resources to advance acoustic monitoring for use in research, management, stewardship, and education.

#### Passive acoustic monitoring can enhance current monitoring efforts by:

- Providing high resolution temporal coverage via continuous sampling or high-frequency intervals (e.g. every 20-60 min.), while leaving recorders deployed for months at a time
- > Operating in poor weather and visibility
- > Utilizing recording systems that are affordable and easily deployed/retrieved

## **Data Collected**

Soundscape metrics which correlate with biodiversity, abundance, and indicate ecosystem function Remote sensing presence & behavior Habitat use: spawning sites, species of interest/ concern Anthropogenic and physical noise Changes in phenology: spawning & reproductive productivity

## Sources of Sound

#### There are three main types of underwater sound<sup>11,12</sup>:

Biological sound is often associated with species-specific behavior such as communication, navigation, foraging, and reproduction. Acoustic monitoring provides a noninvasive, continuous method to monitor animal behavior, habitat quality, habitat use, and community structure over space and time and utilizes sound-producing species as indicators of ecosystem health.

Examples: shrimp, fish, dolphins, manatees, whales, alligators

**Geophysical sound** is sound that is attributed to geophysical processes. Within estuaries, wind, waves, rain, and thunder are common components of the soundscape.

Examples: wind, waves, rain

Anthropogenic sound or human generated noise associated with commercial shipping, recreational boating, dredging, pile-driving, seismic exploration, and energy production have increased dramatically over the last century, and these human activities expose marine organisms to increasing levels of low-frequency noise. This noise can affect marine life by causing hearing threshold shifts, direct physical damage to auditory structures, masking of communication signals, and increased stress levels.

Examples: recreational boats, container ships, dredging, ferries

## **Sound Measurement**

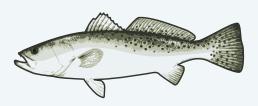


- > Passive acoustics uses hydrophones to detect and measure sound waves, which can be used to identify soniferous species present in an area over space and time. The sounds recorded can be characterized by their loudness (amplitude) and pitch (frequency)13.
- > The most common metric is the acoustic pressure or the sound pressure level (SPL) measured in dB (decibel relative to  $1 \mu Pa$ ).
- Additional metrics include counts of species, number of shrimp snaps, calling intensity, number of vocalizations or noise occurrences
- > Soundscapes in an estuarine environment contain a range of different species of fish and marine mammals vocalizing at different pitches/frequency ranges.
  - Low frequency SPLs may include fish calls from a variety of species, the lower band-

## ACOUSTIC MONITORING // Sound Measurement

width of snapping shrimp snaps, physical sounds, anthropogenic noise, bottlenose dolphin, North Atlantic right whale, and manatee vocalizations.

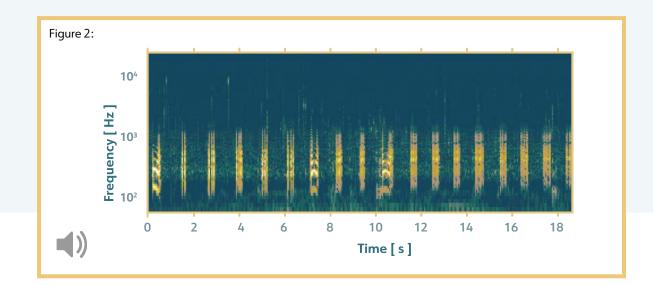
 High frequency SPLs may include bottlenose dolphin vocalizations and the upper bandwidth of snapping shrimp snaps, non-biological sounds such as physical sounds (i.e. waves, rain, wind) or anthropogenic noise (i.e. recreational boats, ships, dredging).



]mmmmmm

## **Spotted Seatrout**

Recording and spectrogram of a Spotted Seatrout *(Cynoscion nebulosus)* vocalization produced during courtship and spawning. The sound contains grunts and staccato sounds centered around 250-500 Hz<sup>5, 14</sup>.



## **Data Collection & Analysis**

The equipment needed to collect passive acoustic data and monitor soundscapes includes a hydrophone and data recording device, which are usually contained within the same housing, and a deployment apparatus. The type and sensitivity of the hydrophone can vary based on the application and intensity of the sounds being measured. The recorded .wav files are analyzed by listening, visually inspecting graphical representations (spectrograms) (Fig.2), and with digital signal processing to identify the sources and intensity of the sounds. Time series of soundscape metrics can be produced to monitor short-term variability and long-term change. A flow chart of the pre-deployment, deployment, retrieval, and analysis process is outlined below.

## 

## ACOUSTIC MONITORING // Data Collection & Analysis

#### **Pre-deployment**

Testing and calibrating the hydrophone for sensitivity and gain, programming duty cycle (e.g. 2 min. recording per hour), and applying ant-fouling material.



#### Deployment

Deployment apparatus can vary by site. Instruments may be attached to PVC anchored in the sediment, contained within an instrument frame, or strapped to a piling or structure.

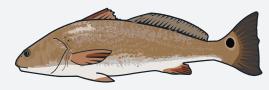
#### Retrieval

Retrieve data logger and dowload data. Audio files are manually reviewed by listening and visually inspecting spectrograms to identify sources of sound using the unique frequency and temporal characteristics. (Figs. 2, 3, 5, 6)



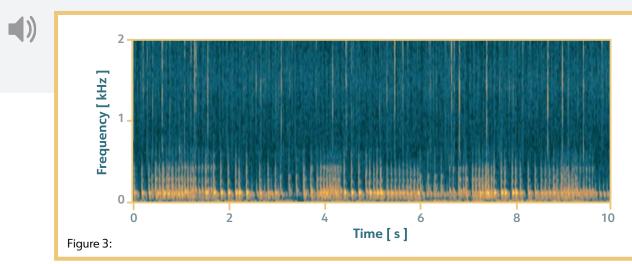
#### Analysis

Temporal patterns of sound production, pressure levels and frequency composition are analyzed (spectral analysis) to produce additional soundscape metrics. (Fig. 4)

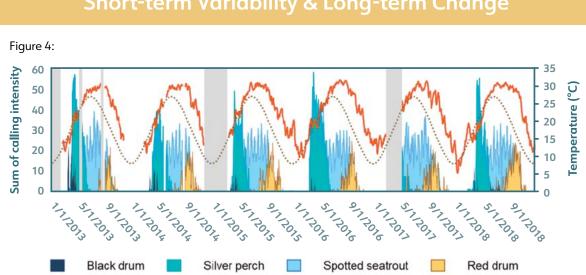


## **Red Drum**

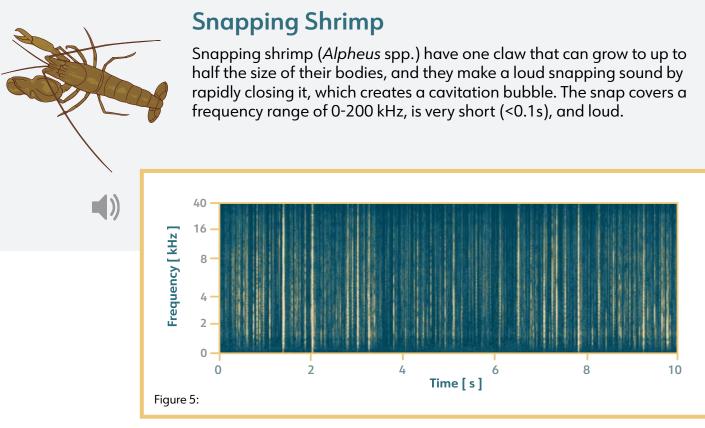
Red Drum (*Sciaenops ocellatus*) vocalization consisting of a low frequency "knock" in the frequency range of 140 - 160 Hz. The number of knocks and pulses of repetition vary among individuals. Sounds are produced by male fish and are associated with spawning<sup>5,15</sup>.



## **ACOUSTIC MONITORING //** Data Collection & Analysis



Time series of courtship and spawning associated vocalizations from Black drum, Silver Perch, Spotted Seatrout, and Red Drum in the May River Estuary, South Carolina, between 2013 and 2018. Acoustic monitoring can be used to identify the onset and duration of spawning seasons and provides a base line from which to assess short-term variability and long-term change<sup>16</sup>.



]mmmmmm

## ;hull[[]]he

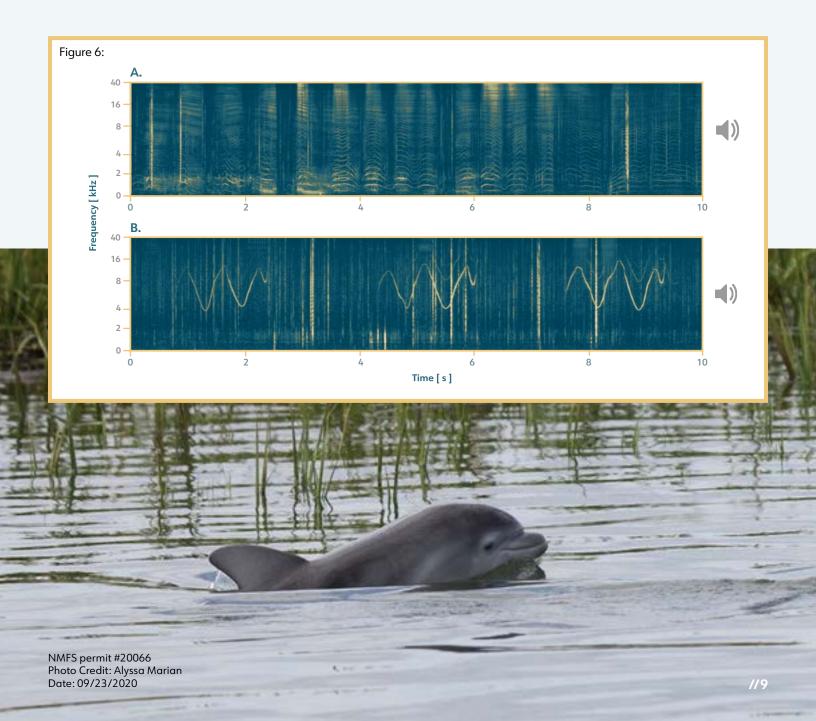
## ACOUSTIC MONITORING // Data Collection & Analysis



## **Common Bottlenose Dolphins**

Two types of vocalizations produced by Bottlenose Dolphins *(Tursiops truncatus)*. A) Burst pulses with harmonic lines, which can vary across a broadband frequency range of 0-160 kHz. and B) Whistles, which range from 2-20 kHz and modulate in frequency.

Echolocation bouts (not shown) are also abundant and the predominant vocalization used for foraging<sup>17</sup>.



ACOUSTIC MONITORING System Wide Monitoring & Habitat Assessment

## The System Wide Monitoring Program of the NERRS

The National Estuarine Research Reserve System (NERRS) is a network of 29 coastal sites designed to protect and study estuarine systems.

Established through the Coastal Zone Management Act, the reserves represent a partnership between NOAA and the coastal states. NOAA provides funding and national guidance, and each site is managed on a daily basis by a lead state agency or university with input from local partners.

Long-term monitoring of meteorological conditions, water quality, and primary production are collected via the System-Wide Monitoring Program (SWMP) at multiple locations within each NERR, with the goal of identifying trends in short-term variability and long-term change in coastal ecosystems. All SWMP data is publicly available and can be <u>downloaded</u>.

Implementing acoustic monitoring at existing SWMP monitoring stations will provide high temporal resolution data of biological, geophysical, and anthropogenic sounds. This approach will provide information on short-term variability and long-term change in higher trophic level organisms beyond what is currently targeted by SWMP.

The three Reserves currently being considered are the Mission-Aransas NERR, Rookery Bay NERR, and North Inlet-Winyah Bay NERR.



Mission-Aransas NERR, Texas



Rookery Bay NERR, Florida

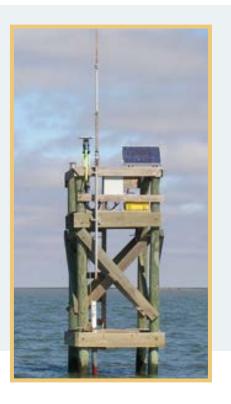


North Inlet-Winyah Bay NERR, South Carolina

## ACOUSTIC MONITORING // SWMP Stations

#### Benefits of hydrophones installed at SWMP stations:

- Passive acoustic monitoring enhances the resolution of biodiversity assessments<sup>4,18</sup>
- Can detect cryptic species that otherwise have gone undetected by visual surveys<sup>19</sup>
- Monitors biological activity at higher trophic levels, which are not currently monitored
- > Useful in estuarine waters with limited visibility
- Understanding when spawning, nesting, and feeding occurs can inform decisions on protecting key habitats and passive acoustics can help provide this information <sup>5, 6, 16, 17</sup>



## Targeted Habitat of Interest (Oyster Reefs)

#### Healthy, Degraded, Restored Oyster Reefs

The ecological functions of oyster reefs feed either directly or indirectly into several critical ecosystem services. These ecosystem services include oyster production, water filtration, carbon sequestration, recreational fisheries production, habitat provision, and habitat and shoreline stabilization<sup>20</sup>.

Transient fishes including red drum, black drum, and spotted seatrout are commonly captured near oyster reefs during community sampling, suggesting that oyster reefs are important foraging areas for predatory fishes<sup>21</sup>.

Sound intensity and diversity is greater on healthy oyster reefs compared to degraded reefs or soft-bottom habitats (Fig.8), and oyster larvae settle in response to reef-associated sounds<sup>2,22</sup>.

Acoustic signatures of oyster reefs convey information about the habitat quality and the organisms that inhabit them. Monitoring the acoustic signatures of natural and restored reefs can help evaluate the rate of recolonization and ecosystem function<sup>23</sup>. Acoustic monitoring also provides biodiversity metrics for these areas, which are difficult to sample using traditional methods (trawls).

## ACOUSTIC MONITORING // Oyster Reefs

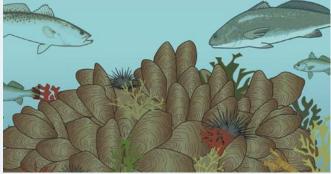
## Signs of a Healthy Reef:

- > High biodiversity across trophic levels
- > High density of oysters
- Numerous resident fish and invertebrates
- > Rich biological sounds associated with foraging, communication, & movement
- > Elevated sound pressure levels

### Signs of a Degraded Reef:

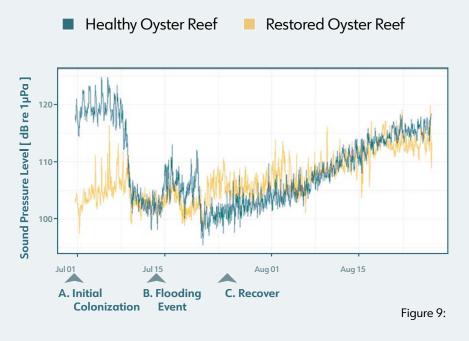
Figure 8:

- Low biodiversity, especially at higher trophic levels
- Dominated by algal species and sediment
- > Diminished sound pressure levels
- Lower SPL and acoustic diversity indices



#### Mission Aransas NERR

Acoustic monitoring of low-frequency (50 - 2500 Hz) sound production (sound pressure level) at an established oyster reef (blue line) and a newly restored oyster reef (orange line) within in the Mission-Aransas NERR. Initially there is a large difference in sound production between the two sites indicating a lack of biological activity at the restored oyster reef (A). Both sites were affected by a flooding event and experienced a decrease in salinity, which corresponded with a cessation of sound production (B) followed by a period of recovery of biological activity and convergence of ecosystem function between the healthy and restored reef (C).



## **STEWARDSHIP** Impacts of Noise and Monitoring Visitor Use

## Impacts of Anthropogenic Noise

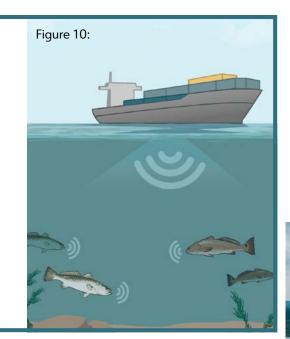
The ocean has become a very noisy environment due to coastal development, commercial shipping, recreational boating, dredging, pile-driving, energy production, and other human activities. Marine organisms are impacted by these activities in a variety of ways including increased stress levels, masking of communication signals, hearing threshold shifts, and damage to auditory structures<sup>7, 8, 24</sup>. Passive acoustics provides a measure of human generated noise and a means to assess potential ecosystem impacts.

Vessel noise varies with the size and speed of ship as well with the type of engine, but generally produces broadband sound that is often loudest within lower frequencies. The high-intensity low-frequency sounds of boat engines coincide with the frequency of biological sound production (Figure 10). Anthropogenic noise may be persistent (e.g active shipping channels), temporary (e.g. areas of dredging) or intermittent (e.g. recreational boat traffic).

#### Impacts of anthropogenic noise

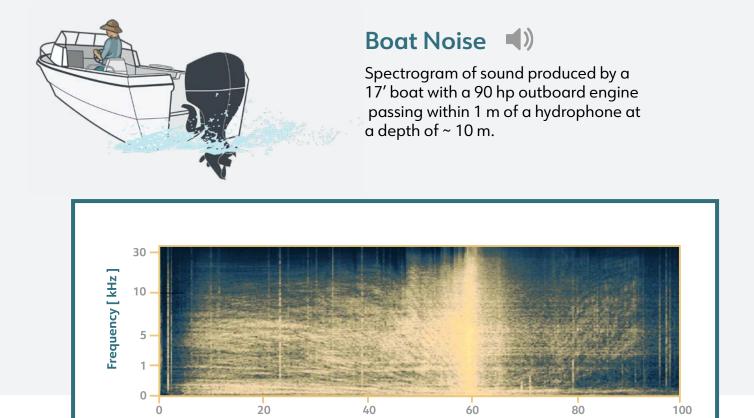
\are largely unstudied in organisms other than marine mammals. However, some of the likely impacts include:

- Disruption of communication through masking (noise at the same frequency of communication)
- Interruption of courtship calls produced by male fish during the spawning process
- > Increased stress hormone levels
- > Alteration of behavior





## **STEWARDSHIP** // Impacts of Noise/Monitoring Visitor Use



Time [s]

Figure 11:

## **Monitoring Visitor Use**

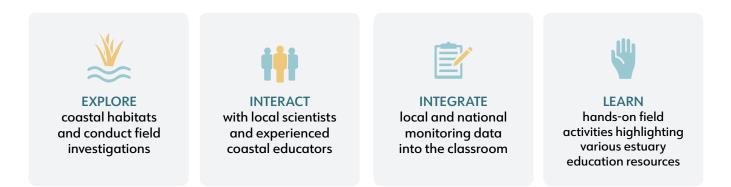
Vessel presence and visitor use are important metrics in assessing ecosystem value and management effectiveness. Acoustic monitoring can provide this information continuously over extended periods of time. Boat noise and other anthropogenic sources of sound can be identified via manual detections or automated processes. The data can be used to measure seasonal, daily, or hourly usage, and duration of visit. Monitoring the level of compliance with regulations may also be of interest as the data collected can indicate when patrols or enforcement measures would be most effective.



**EDUCATION** Teachers on the Estuary (TOTE)

## Teacher Training Workshop

A Teachers on the Estuary (TOTE) workshop is a research and field-based training program held at various research reserve sites. TOTE workshops offer a minimum of 15 contact hours, giving teachers the opportunity to:



Teachers use TOTE to increase their understanding of estuary science, and they learn how to engage students in the investigation of changes in their local environment using data from the NERRS System-Wide Monitoring Program.



## Calling all Spotted Seatrout // Can you hear me?

**Description:** Students will explore the reasons for and challenges to organisms using sound for their survival in an aquatic environment, such as the bays, estuaries, and waterways within the NERR. They will conduct experiments, gather evidence and examine data to better understand the role sound plays in the lives of Spotted Seatrout, and how researchers study the Seatrout sounds to inform the management of their populations.

#### Learning Objectives: Students will...



Explain why and how animals use sound for their survival, considering both land/terrestrial and water/aquatic environments.



Conduct experiments that demonstrate two to three important characteristics of sound, such as amplitude/intensity (loudness), frequency (pitch) and rhythms, and how a water/aquatic environment influences these characteristics.

Identify and predict how human activities might impact Spotted Seatrout communication (and their survival).



Recognize how scientists use specialized equipment, such as hydrophones and acoustic receivers, to capture and measure sounds natural and human-produced in the water/aquatic environment.



Describe why and how scientists study and measure Spotted Seatrout sounds in an estuary.

## **Spotted Seatrout Research**

Scientists have been investigating the sounds that fish make for many years. They study fish sounds to better understand when and where particular fish species are aggregating to spawn and how these aggregations are overlapping with various habitats



and recreational fishing efforts. Spotted Seatrout are of particular interest to these scientists due to their importance as a piscivorous predator, a recreational fishery and the economic importance of the fishery to the regional economy. Scientists have been combining local knowledge sourced from the fishing community with hydroacoustic data to generate spawning aggregation "maps." These maps, along with other research findings, can be used to inform management practices that protect valuable seatrout habitat as well as the fish themselves.



EDUCATION // Student Activities

## **Excercise 1: Animal Sounds**

## > Why do animals make sounds?> How do animals make sounds?

Students will draw on prior knowledge and provided references as they Think/Pair/Share answers to these questions, while learning the basic sound characteristics of pitch/frequency and loudness/intensity as it relates to how we perceive (hear) sound and how we measure sound. They will also learn about the units used to measure sound, decibels (dB) for loudness/ intensity and hertz (Hz) for pitch/frequency.

### **Activity: Sounds Underwater**

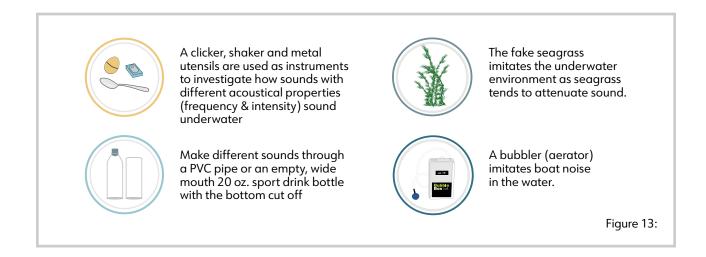
Students will explore sound perception (how we hear sound) in both air and water environments. They will conduct a series of experiments that demonstrate the differences and similarities of sound in different mediums.

#### **Experiment: Exploring Sounds Underwater**

Figure 12:



## EDUCATION // Student Activities



## **Excercise 2: Studying Sounds**

> Why do scientists study animal sounds?> How do scientists study animal sounds?

This exercise introduces the reasons why scientists study animal sounds, and some of the methods they use to listen, record and document animal sounds.

## **Activity: Hearing Sounds**

Students will listen to a variety of animal sounds, as well as other geophysical and anthropogenic sounds generated underwater from a soundscape recording. They will consider what created the sound (what type of animal or what type of activity) and what identifiable characteristics each sound has, such as loudness, pitch and pattern. They will articulate the sound characteristics in writing or illustration.



Play sounds at individual sound stations, or for the whole group.



Aid students in their identification of the sounds by providing them with a set of images of the animal or machine/activity to choose from and match up.





Listening through headphones will provide the best listening experience, especially to limit background noise and to focus their attention.



After listening to recordings, students should compare their identifications (what/ who made the sound) and their sound descriptions.



Some people can hear a wider range of sounds than others and our description of sounds is solely based on our perception and interpretation of the sound. The sound source (who/what) discussion can be extended to include a consensus making component where all the students work together to identify who/what made the sound.

## **Activity: Seeing Sounds**

Students will observe sound recording spectrograms, which illustrate the frequency and intensity of sound through time. Students will use what they know about sounds, including their own sound notes/illustrations, to match up the sound source (who/what made the sound) with the spectrogram.

- > What do you notice about the graph?
- Can you identify the units of measurement? Is it decibels or hertz? What does this tell you about the sound?
- > Which animal or human activity/machine made the sounds graph you are looking at? Explain your reasoning.
- > How did your description of the sound compare with the graph? How did it differ?

## **Exercise 3: Spotted Seatrout Case Study**

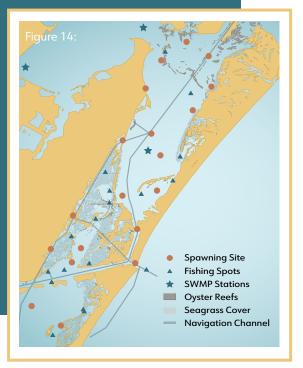
Students will be introduced to a specific research project being conducted at a NERR. Students will engage in an exercise that highlights how the scientists used sound to answer their research question.

### **Activity: Monitoring Sounds**

Scientists were interested in learning more about Spotted Seatrout spawning aggregations within an estuary, specifically, where they are located, when they occur and how fishing activities overlap or coincide with these events. This knowledge can be very valuable to scientists that seek to understand recreational fishery populations and provide insight on how to conserve them for future generations. Knowing seatrout make sound to communicate during spawning, scientists randomly sampled sites in the estuary to see where spawning was occurring and then compared those sites with known fishing locations.

#### Students will observe and interpret maps illustrating "prime fishing locations" as well as known habitat locations, and spawning sites.

- > Observe the map of sea trout spawning locations. Determine where human activities might impact or overlap with sea trout. What locations are most at risk of noise pollution or interference? Why do you think that?
- > How might the noise interference impact the seatrout communication?





## **REFERENCES // RESOURCES**

- 1. Mueller C, Monczak A, Soueidan J, McKinney B, Smott S, Mills T, Ji Y, Montie EW. 2020. Sound characterization and fine-scale spatial mapping of an estuarine soundscape in the southeastern USA. *Marine Ecology Progress Series* 645:1-23.
- 2. Bohnenstiehl DR, Lillis A, Eggleston DB. 2016. Shrimp Sound in Sub-Tidal Oyster Reef Habitat. *PLoS ONE* 11:143691.DOI: 10.1371/journal.pone.0143691.
- **3.** Montie EW, Vega S, Powell M. 2015. Seasonal and spatial patterns of fish sound production in the May River, South Carolina. *Transactions of the American Fisheries Society* 144:705–716. DOI: 10.1080/00028487.2015.1037014.
- 4. Monczak A, McKinney B, Mueller C, Montie EW. 2020. What's all that racket! Soundscapes, phenology, and biodiversity in estuaries. *PLoS ONE* 15:1–18. DOI: 10.1371/journal.pone.0236874.
- 5. Monczak A, Berry A, Kehrer C, Montie EW. 2017. Long-term acoustic monitoring of fish calling provides baseline estimates of reproductive time-lines in the May River estuary, southeastern USA. *Marine Ecology Progress Series* 581:1–19. DOI: 10.3354/meps12322.
- 6. Biggs CR, Lowerre-Barbieri S, Erisman B. 2018. Reproductive resilience of an estuarine fish in the eye of a hurricane. *Biology Letters* 14:10-14.
- 7. Smott S, Monczak A, Miller M, Montie EW. 2018. Boat noise in an estuarine soundscape a potential risk on the acoustic communication and reproduction of soniferous fish in the May River, South Carolina. *Marine Pollution Bulletin* 133, 246-260.
- 8. Slabbekoorn H, Bouton N, van Opzeeland I, Coers A, ten Cate C, Popper AN. 2010. A noisy spring: the impact of globally rising underwater sound levels on fish. *Trends in Ecology & Evolution* 25: 419-27.
- Indeck KL, Simard P, Gowans S, Lowerre-Barbieri S, Mann DA. 2015. A severe red tide (Tampa Bay, 2005) causes an anomalous decrease in biological sound. *Royal Society Open Science* 2(9). DOI: 10.1098/rsos.150337.
- 10. Rossi T, Connell SD, Nagelkerken I. 2016. Silent oceans: ocean acidification impoverishes natural soundscapes by altering sound production of the world's noisiest marine invertebrate. *Proceedings of the Royal Society* B 283:20153046. DOI: 10.1098/rspb.2015.3046.
- Pijanowski BC, Villanueva-Rivera LJ, Dumyahn SL, Farina A, Krause BL, Napoletano BM, Gage SH, Pieretti N. 2011. Soundscape ecology: The science of sound in the landscape. *BioScience* 61:203–216. DOI: 10.1525/bio.2011.61.3.6.
- 12. Duarte CM, Chapuis L, Collin SP, Costa DP, Devassy RP, Eguiluz VM, Erbe C, Gordon TAC, Halpern BS, Harding HR, Havlik MN, Meekan M, Merchant ND, Miksis-Olds JL, Parsons M, Predragovic M, Radford AN, Radford CA, Simpson SD, Slabbekoorn H, Staaterman E, Van Opzeeland IC, Winderen J, Zhang X, Juanes F. 2021. The soundscape of the Anthropocene ocean. *Science* 371. DOI: 10.1126/science.aba4658.
- **13.** Au WWL, Hastings MC. 2008. Principles of Marine Bioacoustics Modern Acoustics and Signal Processing. New York, NY: Springer.
- 14. Montie EW, Hoover M, Kehrer C, Yost J, Brenkert K, O'Donnell T, Denson MR. 2017. Acoustic monitoring indicates a correlation between calling and spawning in captive spotted seatrout (*Cynoscion nebulosus*). *PeerJ* 5:e2944. DOI: 10.7717/peerj.2944.
- **15.** Montie EW, Kehrer C, Yost J, Brenkert K, O'Donnell T, Denson MR. 2016. Long-term monitoring of captive red drum Sciaenops ocellatus reveals that calling incidence and structure correlate with egg deposition. *Journal of Fish Biology* 88:1776–1795. DOI: 10.1111/jfb.12938.

- **16.** Monczak A, McKinney B, Soueidan J, Marian A, Seder A, May E, Morgenstern T, Roumillat W, Montie EW. 2021. Sciaenid courtship sounds correlate with juvenile appearance and abundance in the May River, South Carolina. Marine Ecology Progress Series. *In Review.*
- **17.** Marian AD, Monczak A, Balmer BC, Hart LB, Soueidan J, Montie EW. 2021. Long-term passive acoustics to assess spatial and temporal vocalization patterns of Atlantic common bottlenose dolphins *(Tursiops truncatus)* in the May River estuary, South Carolina. *Marine Mammal Science*, 37, 1060–1084.
- **18.** Staaterman E, Ogburn MB, Altieri AH, Brandl SJ, Whippo R, Seemann J, Goodison M, Duffy JE. 2017. Bioacoustic measurements complement visual biodiversity surveys: Preliminary evidence from four shallow marine habitats. *Marine Ecology Progress Series* 575:207–215. DOI: 10.3354/meps12188.
- **19.** Picciulin M, Kéver L, Parmentier E, Bolgan M. 2019. Listening to the unseen: Passive acoustic monitoring reveals the presence of a cryptic fish species. *Aquatic Conservation: Marine and Freshwater Ecosystems* 29:202–210. DOI: 10.1002/aqc.2973.
- **20.** Grabowski JH, Brumbaugh RD, Conrad RF, Keeler AG, Opaluch JJ, Peterson CH, Piehler MF, Powers SP, Smyth AR. 2012. Economic valuation of ecosystem services provided by oyster reefs. *BioScience* 62:900–909. DOI: 10.1525/bio.2012.62.10.10.
- 21. Tinhan T, Erisman B, Aburto-Oropeza O, Weaver A, Vázquez-Arce D, Lowe CG. 2014. Residency and seasonal movements in Lutjanus argentiventris and Mycteroperca rosacea at Los Islotes Reserve, Gulf of California. *Marine Ecology Progress Series* 501:191–206. DOI: 10.3354/meps10711.
- **22.** Lillis A, Eggleston DB, Bohnenstiehl DR. 2013. Oyster larvae settle in response to habitat-associated underwater sounds. *PLoS ONE* 8:21–23. DOI: 10.1371/journal.pone.0079337.
- **23.** Zenil P, Encomio VG. 2011. Passive acoustics as a monitoring tool for evaluating oyster reef restoration. *The Journal of Acoustical Society of America* 129.
- 24. Merchant ND, Pirotta E, Barton TR, Thompson PM. 2014. Monitoring ship noise to assess the impact of coastal developments on marine mammals. *Marine Pollution Bulletin* 78, 85–95.

### **Additional Resources**

Biggs CR, Erisman BE. 2021. Transmission loss of Fish Spawning Vocalizations and the Detection Range of Passive Acoustic Sampling in Very Shallow Estuarine Environments. Estuaries and Coasts 44:2026–2038. DOI: 10.1007/s12237-021-00914-5

Bohnenstiehl DR, Lyon RP, Caretti ON, Ricci SW, Eggleston DB. 2018. Investigating the utility of ecoacoustic metrics in marine soundscapes. Journal of Ecoacoustics 2(2):1-1.

Coquereau L, Grall J, Clavier J, Jolivet A, Chauvaud L. 2016. Acoustic behaviours of large crustaceans in NE Atlantic coastal habitats. Aquatic Biology 25:151–163. DOI: 10.3354/ab00665.

Eggleston DB, Lillis A, Bohnenstiehl DR. 2016. Soundscapes and larval settlement: larval bivalve responses to habitat-associated underwater sounds. Advances in Experimental Medicine and Biology 875:255–63.

Lindseth AV, Lobel PS. 2018. Underwater soundscape monitoring and fish bioacoustics: a review. Fishes 3(3).

Lowerre-Barbieri SK, Barbieri LR, Flanders JR, Woodward AG, Cotton CF, Knowlton MK. 2008. Use of passive acoustics to determine red drum spawning in Georgia waters." Transactions of the American Fisheries Society 137(2):562–75.

Merchant ND, Fristrup KM, Johnson MP, Tyack PL, Witt MJ, Blondel P, Parks SE. 2015. Measuring acoustic habitats. Methods in Ecology and Evolution 6:257–265. DOI: 10.1111/2041-210X.12330.

Monczak A, Mueller C, Miller ME, Ji Y, Borgianini SA, Montie EW. 2019. Sound patterns of snapping shrimp, fish, and dolphins in an estuarine soundscape of the southeastern USA. Marine Ecology Progress Series 609:49–68. DOI: 10.3354/meps12813.

National Estuarine Research Reserve. NOAA Office for Coastal Management. coast.noaa.gov/nerrs/.



## RESOURCES

## Authorship

**Christopher R. Biggs –** University of Texas at Austin Marine Science Institute, cbiggs@utexas.edu **Eric W. Montie –** University of South Carolina, Beaufort, emontie@uscb.edu

Matthew E. Kimball - University of South Carolina, matt@baruch.sc.edu

**Robert P. Dunn –** North Inlet-Winyah Bay National Estuarine Research Reserve / University of South Carolina, robert@baruch.sc.edu

Kevin M. Boswell – Florida International University, kevin.boswell@fiu.edu

Lindsey Transue - College of Charleston, South Carolina, transuelm@g.cofc.edu

## Illustration and design

Katie Sedivec - Golden Grouper Creative, https://goldengrouper.com/

## Citation

Biggs, CR, Montie, EW, Kimball, M, Dunn, R, Boswell K, Transue L. 2021. Listen In: Acoustic Monitoring of Estuarine Communities Facing Ecosystem Change. National Estuarine Research Reserve System Science Collaborative. P 23.

